LETTERS TO NATURE

concerned. The strikingly derived nature of the Neanderthal labyrinthine features, already present before birth, and the apparent lack of continuity with modern human morphology could be seen as an argument in support of distinguishing between Neanderthals and modern humans at the species level.

The evidence from Arcy indicates that advanced Châtelperronian industries were used by late Neanderthals, suggesting a high degree of acculturation. The association of the Arcy Neanderthal with personal ornaments so similar to those found in contemporary and nearby Aurignacian layers^{14,29} questions the nature of the cultural interactions with modern humans. At least in the case of these specific objects, we may be facing evidence of a trading process rather than the result of technical imitation of modern human technology by Neanderthals.

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- 1. Demars, P. Y. & Hublin, J. J. ERAUL Liege 34, 29-42 (1989).
- Harrold, F. B. in *The Human Revolution* (eds Mellars, P. A. & Stringer, C. B.) 677–713 (Edinburgh Unviersity Press, Edinburgh, 1989).

 3. Mellars, P. A. Phil. Trans, R. Soc. Lond. B **337**, 225–234 (1992).
- 4. Pelegrin J. Technologie Lithique: le Chatelperronien de Roc-de-Combe (Lot) et de la Côte (Dordogne) (Paris, CNRS Cahiers du quaternaire, 1995). 5. Straus, L. G. Evol. Antrhrop. 2, 195–198 (1993–1994).
- Clark, G. A. & Lindly, J. M. in The Human Revolution (eds Mellars, P. A. & Stringer, C. B.) 621-
- 676 (Edinburgh University Press, Edinburgh, 1989). 7. Wolpoff, M. & Frayer, D. *Nature* **356,** 200–201 (1992).
- 8. Stringer, C. & Grün, R. *Nature* **351**, 701-702 (1991). 9. Léveque, F. & Vandermeersch, B. C.R. Acad. Sc. Paris **291**, 187–189 (1980). 10. Mercier, N. et al. *Nature* **351**, 735-739 (1991).
- 11. Leroyer, C. & Leroi-Gourhan, A. Bull. Soc. Prehist. Fr. **80,** 41–44 (1983). 12. Leroyer, C. ERAUL Liege **35.** 103–108 (1988).
- 13. Baffier, D. & Julien, M. in Paléolithique Moyen Recent et Paléolithique Superieur Ancien en Europe (ed. Fairizy, C.) 329-334 (Musée de Préhistoire d'Ile de France, Nemours, 1990)

- 14. Taborin, Y. in Paléolithique Moyen Recent et Paléolithique Superieur Ancien en Europe (ed. Fairizy, C.) 335–244 (Musée de Préhistoire d'Ile de France, Nemours, 1990). 15. Leroi-Gourhan, A. *Ann. Paléont*, **44**, 87–148 (1958).
- 16. Hedges, R. E. M., Housley, R. A., Bronk Ramsey, C. & Van Klinken, G. J. Archaeometry 36, 337-374 (1994)
- 17. Girard, M., Miskovsky, J. C. & Evin, J. in Paléolithique Moyen Recent et Paléolithique Superieur Ancien en Europe (ed. Fairizy, C.) 295–303 (Musée de Préhistoire d'Ile de France, Nemours, 1990)
- 18. Spoor, F., Wood, B. & Zonneveld, F. Nature **369**, 645–648 (1994)
- Spoor, C. F. & Zonneveld, F. W. Cour. Forsch. Senckenberg 171, 251–256 (1994).
 Spoor, C. F. & Zonneveld, F. W. J. Anat. 186, 271–286 (1995).
- 21. Spoor, C. F. The Comparative Morphology and Phylogeny of the Human Bony Labyrinth (Utrecht University, Utrecht, 1993).

 22. Ruff, C. B. *Yb. of Phys. Antrhrop.* **37.** 65–107 (1994).
- 23. Hublin, J. J., Barroso Ruiz, C., Medina Lara, P., Fontugne, M. & Reyss, J.-L. C.R. Acad. Sc. Paris 321 Ila, 931-937 (1995).
- 24. Bischoff, J. L., Soler, N., Maroto, J. & Julia, R. J. Archaeol. Sci. 16, 553–576 (1989). 25. Cabrera-Valdes, V. & Bischoff, J. L. J. Archeol. Sci. 16, 577-584 (1989).
- 26. Bischoff, J. L. et al. J. Archaeol, Sci. 21, 541-551 (1994).
- 27. Howell, F. C. in Origins of Anatomically Modern Humans (eds Nitecki, M. H. & Nitecki, D. V.) 253-319 (Plenum, New York, 1994).
- 28. Rak, Y. in Species, Species Concepts and Primate Evolution (eds Kimbel, W. H. & Martin, L. B.) 523-536 (Plenum, New York, 1994).
- 29. Leujeune, M. L'art Mobilier Paléolithique et Mésolithique de Belgique (Artefacts 4) (Arc, Viroinval, 1987).
- 30. Van Vark, G. N. in Multivariate Statistical Methods in Physical Anthropology (eds Van Vark, G. N. & Howells, W. W.) 323-349 (Riedel, Dordrecht, 1984).

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Continental breakup and the ordinal diversification of birds and mammals

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THE classical hypothesis for the diversification of birds and mammals proposes that most of the orders diverged rapidly in adaptive radiations after the Cretaceous/Tertiary (K/T) extinction event 65 million years ago 1-3. Evidence is provided by the near-absence of fossils representing modern orders before the K/ T boundary^{4,5}. However, fossil-based estimates of divergence time are known to be conservative because of sampling biases⁶, and some molecular/time estimates point to earlier divergences among orders⁷⁻¹⁰. In an attempt to resolve this controversy, we have estimated times of divergence among avian and mammalian orders with a comprehensive set of genes that exhibit a constant rate of substitution. Here we report molecular estimates of divergence times that average about 50-90% earlier than those predicted by the classical hypothesis, and show that the timing of these divergences coincides with the Mesozoic fragmentation of emergent land areas. This suggests that continental breakup may have been an important mechanism in the ordinal diversification of birds and mammals.

Molecular time estimation of evolutionary divergence requires genes that are evolving at a relatively constant rate, which often limits the number of sequences available for analysis^{8,9}. Fortunately, the widespread use of model organisms for genetic FIG. 1 Relationship between the evolutionary histories of mammals and > birds and events in earth history during the Mesozoic and Cenozoic. a, Fossil record of the orders of eutherian mammals and birds⁴ (solid line, certain; broken line, uncertain); avian taxonomy from ref. 29. b, Molecular estimates of divergence times based on constant-rate genes (from Table 1); the number of genes (constant rate/total) is in parentheses; filled circles, mean; horizontal bar, ±s.e.; thin vertical line, median. c, Land breakup (fission) and fusion events determined from reconstructions of emergent land areas during the last 250 Myr (ref. 30); only emergent land areas $>3\times10^6\,\text{km}^2$ (about the size of India) are considered here, although the same pattern is evident if smaller areas (say, the area of Madagascar) are considered. d, Total emergent land area exposed at different times during the last 250 Myr (ref. 30).

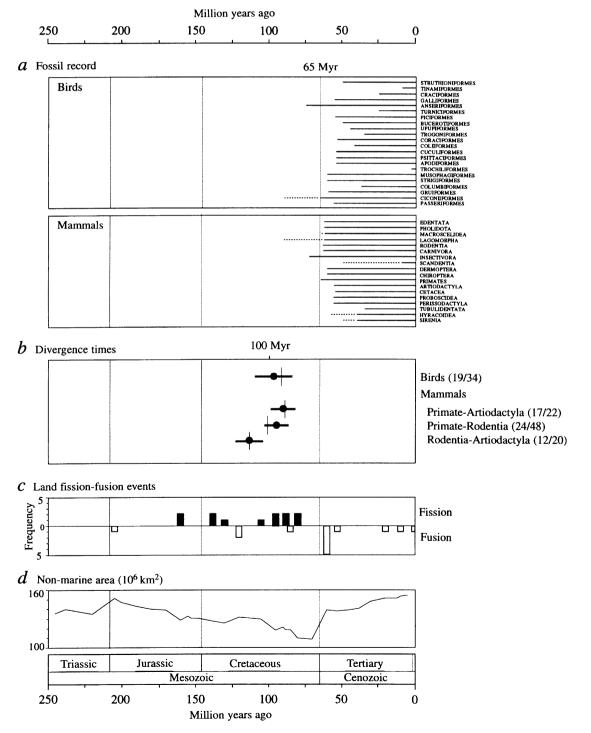
METHODS. For avian comparisons (in b), the nuclear genes used and taxa are (g, galliform; a, anseriform; c, columbiform; s, struthioniform): acylphosphatase-muscle_{ga}, alcohol dehydrogenase_{gs}, α -crystallin_{gacs}, α -globin_{gacs}, argininosuccinate lyase_{ac}, β2 thyroid hormone receptor_{hm}, β-globin_{gacs}, cytochrome c_{gacs} , embryonic α -globin $_{\text{ga}}$, histone $\mathbf{1}_{\text{ga}}$, lactate dehydrogenase \vec{B}_{ga} , lysozyme \vec{c}_{ga} , malic enzyme_{ac}, MX protein_{ga}, prolactin receptor_{gc}, and somatotropin_{ga} (for all genes, sequences of a mammal and an outgroup (or paralogous sequence) also were used). For mammalian comparisons, the nuclear genes used and taxa are (h, H. sapiens; m, M. musculus; b, B. taurus): acetylcholine receptor $\alpha_{\text{hmb}}\text{, acetylcholine receptor }\delta_{\text{hmb}}\text{, acetyl-}$ choline receptor γ_{hm} , α -globin_{hmb}, annexin II_{hm}, Bc12-Ig fusion gene_{hm}, B myb_{hm} , casein kinase II α_{hb} , CD18-integrin β -2_{mb}, CDC2_{hmb}, c-kit proto $oncogene_{hm}$, C-myb_{hmb}, connexin 43_{hmb} , cyclin A_{hmb}, cyclophilin B_{hb}, c-yes protein_{hm}, E-cadherin_{hm}, erythroblast virus oncogene homologue 2_{hm}, fattyacid-binding protein $_{hmb}$, focal adhesion kinase $_{hm}$, follistatin $_{hmb}$, GATA- 3_{hm} , histone H2A.X_{hm}, lactate dehydrogenase A_{hmh}, lamin B1_{hm}, midkine pro- $\mathsf{tein}_{\mathsf{hm}}$, MOS proto-oncogene $_{\mathsf{hm}}$, muscle pyruvate kinase $_{\mathsf{hm}}$, myelin proteolipid protein_{hmb}, Myf- 5_{hm} , Na⁺/K⁺ ATPase β - 2_{hm} , NAD⁺ ribosyltransferase_{hmb}, N-cadherin_{hm}, neural cell adhesion molecule_{hmb}, neu $rotrophin-3_{hm},\ NF-\kappa-B_{hm},\ N-myc_{hm},\ nucleolar\ protein\ NO38_{hm},\ nucleolin_{hm},$ ornithine decarboxylase_{hmb}, p53 cellular tumour antigen_{hmb}, S6 II kinase_{hm}, $SPARC_{hmb}\text{, transcription factor Eryf-}1_{hm}\text{, }TGF-\beta2_{hm}\text{, }TGF-\beta3_{hm}\text{, transglutami-}$ nase_{hmb}, tyrosine phosphatase_{hm}, vimentin_{hmb} (for all genes, sequences of G. gallus and an outgroup, usually X. laevis or a paralogous sequence, also were used).

research has generated a large number of gene sequences for representatives of several orders of birds and mammals. This now allows us to derive a single average estimate of divergence time from many independent (single gene) estimates, thus reducing potential biases that may exist with individual genes, and increasing the accuracy of the time estimate.

An accurate calibration point is also required for a reliable estimate of divergence time from molecular data. Typically, the first occurrence of two taxa in the fossil record is used as a minimum estimate of their time of divergence. However, that may be a considerable underestimate (> 50%) because of sampling biases⁶. The fossil record of birds and mammals is particularly prone to such biases because of the long time span between the earliest fossils (150 million years (Myr) for birds, 220 Myr for

mammals) and the first appearances of the modern orders (mostly 55–65 Myr)⁴. Therefore, an internal calibration based on the fossil record of the modern orders themselves may lead to a large bias in the divergence times estimated. To avoid this, we have chosen the relatively well-constrained fossil divergence time between the ancestors of birds (diapsid reptiles) and mammals (synapsid reptiles), 310 Myr, as an external calibration point⁴. The first appearance of amniotes at 335 Myr (ref. 11) and of tetrapods at 370 Myr (ref. 12) suggests that a divergence time of birds and mammals much earlier than 310 Myr is unlikely.

All of our molecular estimates of divergence times for avian and mammalian orders are Mesozoic rather than Cenozoic, and are considerably older than divergence times suggested by fossil evidence (Table 1; Fig. 1). Some previous estimates of divergence



times based on immunological distances and sequence data also pre-date the K/T boundary⁷⁻¹⁰. However, unlike those studies, we have tested for rate constancy both among orders and between the lineages used for calibration, used an external (bird-mammal) rather than an internal calibration, and based our divergence times exclusively on a large number of constant-rate genes.

We suggest that the fragmentation of emergent land areas during the Cretaceous, not the sudden availability of ecological niches following the K/T extinction event, was the mechanism responsible for the diversification of avian and mammalian orders. When plate tectonics and sea level are considered together, all of the major continental breakup events during the last 250 Myr occurred in the Mesozoic, and mainly during the Cretaceous (Figs 1c and 2). The timing of these events corresponds closely to the molecular time estimates for divergences of the orders (Fig. 1b). The tectonic breakup of Pangaea has been cited as a factor in the historical biogeography of some Mesozoic organisms, includbirds and mammals^{9,13}. However, few data on times of divergence from constant-rate genes have been available to test such a hypothesis. Moreover, comparisons have not been made between ordinal divergence times of multiple groups such as birds and mammals. The continental breakup hypothesis (Fig. 2) is compatible with the extensive speciation that occurred within orders following the sudden availability of niches in the early Tertiary period.

These early molecular dates for divergences among the orders of birds and mammals indicate an unusually strong bias in the fossil record against Middle to Late Cretaceous fossils. Other groups of terrestrial vertebrates, including dinosaurs, also show a mid-Cretaceous decline in diversity¹⁴, but not as pronounced. Besides the sampling bias already noted⁶,

the highest sea levels in the Mesozoic and Cenozoic occurred in the Middle to Late Cretaceous, resulting in about 25% less emergent land area than exists at present (Fig. 1d). This would have proportionately reduced the area available for the deposition of terrestrial fossils.

The general concordance between ordinal diversification and Earth history for birds and mammals predicts that similar patterns may be seen in other terrestrial organisms. Although the relationships and times of divergence of turtle, lizard and snake families are not yet well established^{4,15}, Mesozoic continental fragmenta-

TABLE 1 Molecular time estimates for the divergence of avian and mammalian orders*

Comparisons	No Total	o. of genes Constant rate	Total length†	Time‡ (Myr)	Evolutionary rate $(\times 10^{-4} \text{ per site per Myr})$
Birds Nuclear genes§ Mitochondrial genes	34	19	2,829	$\textbf{97} \pm \textbf{12}$	1.9–13.1
rRNA Protein	2 3	2 0	1,267	68–131	2.8
Mammals Nuclear genes					
Primate-Rodent	48	24	10,051	95 ± 7	0.8-8.5
Primate-Artiodactyl	22	17	6,573	90 ± 8	0.4-10.0
Rodent–Artiodactyl Mitochondrial genes	20	12	4,497	$\textbf{113} \pm \textbf{9}$	1.0-10.2
rRNA∥ Protein	2 13	2 0	2,171	99–184	4.0

Nuclear genes. Complete amino-acid sequences (≥ 100 residues) of nuclear genes were obtained (HOVERGEN18 3, release 10; ENTREZ, release 18). Avian taxa included a struthioniform (Struthio camelus), galliform (Gallus gallus), anseriform (Anas platyrhynchos or Cairina moschata) and columbiform (Columba livia); corresponding sequences of a mammal (for calibration) and a more distantly related taxon (usually Xenopus laevis or a paralogous sequence, for root) were included. Mammalian taxa included a primate (Homo sapiens), rodent (Mus musculus) and artiodactyl (Bos taurus); relationships among these orders generally have not been well established¹⁹, although molecular evidence has favoured a basal position for rodents¹⁰. For mammalian analyses, each comparison included data from a bird (G. gallus) for calibration and an outgroup (usually X. laevis or a paralogous sequence) for root. Sequences of individual genes (or gene families) were aligned²⁰ and orthology determined by phylogenetic analysis using MEGA²¹. Sites with alignment gaps were removed and constancy of rate between mammalian orders and between G. gallus and the mammals was tested (5% level)²². Pairwise distances were estimated by use of a gamma correction²³ with shape parameter (a) of 2. For each gene, average pairwise distance between G. gallus and the two mammalian sequences was divided by 620 (2 \times 310 Myr) to obtain evolutionary rate, and the pairwise distance between mammalian orders was converted to time using this estimate of rate. Ordinal divergences were estimated with the Poisson correction distance and a different gamma distance (a = 1); resulting estimates were within $\pm 5\%$ of those in the table. Assuming a basal position of rodents, the average time estimate (nuclear genes) of the divergence of the rodent lineage from the primate-artiodactyl lineage is 104 Myr. The timing of avian ordinal divergences was accomplished similarly, although fewer sequences (≥ approximately 100 residues) were available, so all pairwise comparisons for nuclear genes were pooled to estimate means. S.e.m. was computed by dividing the standard deviation by the square root of the number of different genes to account for non-independence in multiple comparisons. Mitochondrial genes. For avian comparisons, portions of the two rRNA genes were analysed by the linearized tree method using a gamma correction distance (a = 1.7, transversional distance under Kimura's model²¹) for representatives of 7 avian orders: galliform (G. gallus), anseriform (Anas platyrhynchos), struthioniform (Rhea americana), psittaciform (Melopsittacus undulatus), columbiform (Columbia livia), cuculiform (Cuculus pallidus), and ciconiiform (Ciconia nigra). The amino-acid sequences of ATP6, ATP8 and COXII from G. gallus and A. platyrhynchos²⁴ also were analysed using a gamma correction distance (a = 1.4, Poisson model²¹). In both cases, an artiodactyl (B. taurus) and carnivore (Phoca vitulina) were used for calibration, and a frog (X. laevis) was included for root. For mammalian comparisons, the taxa²⁵ are: an artiodactyl (B. taurus), carnivore (P. vitulina), cetacean (Balaenoptera physalus), perissodactyl (Equus caballus), primate (H. sapiens), rodent (M. musculus), a galliform bird (G. gallus, for calibration), and a frog (X. laevis, for root). The linearized tree method²⁶ was used with the gamma distance (a = 1.4 for protein genes, a=1.7 for rRNA genes)^{21,27}. In both avian and mammalian analyses using linearized tree methods, rate constancy was rejected for protein genes but not for rRNA genes. However, the likelihood ratio tests²⁸ (using Kimura's model with R=2) rejected rate constancy in rRNA genes as well. Therefore dates obtained by the linearized tree method are not shown in Fig. 1.

- * Nuclear genes were analysed individually, and mitochondrial genes were analysed tandemly as two datasets (protein and rRNA genes). Total sequence lengths, times of divergence, and evolutionary rates are shown only for genes in which constancy of evolutionary rate was not rejected.
 - † Number of aligned amino-acid residues (proteins) or nucleotides (rRNA) for constant-rate genes.
- \ddagger Mean time of divergence ($\pm 1\,\mathrm{s.e.}$) as computed from the time estimates for comparisons of nuclear genes. For mitochondrial genes, the range of time is given for all ordinal divergences estimated in the linearized trees.
 - § Based on pairwise comparisons of galliform, anseriform, columbiform and struthioniform birds.
 - || Constant rate was rejected in likelihood ratio tests but not in linearized tree tests.

tion has been implicated in dinosaur biogeography¹⁶. Similarly, recent DNA sequence evidence about frog relationships¹⁷ identified two major groups of families associated with Laurasia (Archaeobatrachia) and Gondwana (Neobatrachia); the two major neobatrachian groups are associated with South America (Bufonoidea) and Africa (Ranoidea). These observations conform to the hypothesis of continental breakup and diversification. Major radiations of marine invertebrates in the mid-Cretaceous also have been causally linked to the subdivision of continents by epicontinental seas¹³.

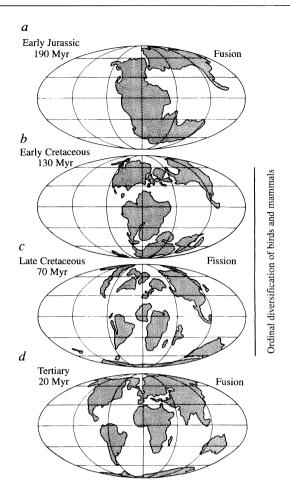


FIG. 2 Breakup (fission) and fusion of emergent land areas in the Mesozoic and Cenozoic as determined by plate-tectonic reconstructions and paleocoastline data (modified from ref. 30). a, Early Jurassic (190 Myr), with 2 large continents. b, Early Cretaceous (130 Myr), with 7 continents $(> 3 \times 10^6 \text{ km}^2)$, c, Late Cretaceous (70 Myr), with 11 continents. d, Mid-Tertiary (20 Myr), with 5 continents; subsequent fusion of North and South America, and Africa with Eurasia, led to the present configuration of three isolated emergent land masses. The orders of mammals and birds diverged during the Cretaceous (for example, b and c) at the time when emergent land areas were undergoing breakup.

If continental breakup were an important mechanism for ordinal diversification in birds and mammals, then it might be expected that distributional data of living fossil taxa should help in biogeographic reconstruction. Although some orders can be associated with specific palaeolandmasses⁹, most now occur on multiple continents and their original distributions have not been established. This is not surprising considering that a large amount of continental fusion, allowing migration, already had occurred by the Early Tertiary when the fossils of most modern orders first appeared (Fig. 1a, c). As larger numbers of constant-rate genes for a diversity of taxa become available, it should be possible more accurately to associate ordinal divergences with specific continental breakup events. \Box

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- 1. Romer, A. S. Vertebrate Paleontology (Univ. Chicago Press, 1966).
- Strickberger, M. W. Evolution (Jones and Bartlett, Boston, MA, 1995).
- Feduccia, A. Science **267**, 637–638 (1995).
 Benton, M. J. (ed.) *The Fossil Record* Vol. 2 (Chapman and Hall, New York, 1993).
- Chiappe, L. M. Nature **378**, 349-355 (1995).
- 6. Martin, R. D. Nature 363, 223-234 (1993). 7. Prager, E. M., Brush, A. H., Nolan, R. A., Nakanishi, M. & Wilson, A. C. J. molec. Evol. 3, 243-
- 8. Li, W.-H., Gouy, M., Sharp, P. M., O'Huigin, C. & Yang, Y.-W. Proc. natn. Acad. Sci. U.S.A. 87,
- 9. Easteal, S., Collet, C. & Betty, D. The Mammalian Molecular Clock (Landes, Austin, TX, 1995).

- 10. Janke, A., Feldmaier-Fuchs, G., Thomas, W. K., von Haeseler, A. & Pääbo, S. Genetics 137, 243-256 (1994)
- Smithson, T. R. Nature **342**, 676–678 (1989).
- 12. Ahlberg, P. E. & Milner, A. R. Nature 368, 507-514 (1994).
- 13. Hallam, A. An Outline of Phanerozoic Biogeography (Oxford Univ. Press, 1994).
 14. Padian, K. & Clemens, W. A. in Phanerozoic Diversity Patterns (ed. Valentine, J. W.) 41–96
- (Princeton Univ. Press, NJ, 1985). 15. Zug. G. R. Herpetology (Academic, San Diego, CA, 1993).
- 16. Russell, D. A. Hist. Biol. **10**, 3–12 (1995).
- Hay, J. M., Ruvinsky, I., Hedges, S. B. & Maxson, L. R. Molec. Biol. Evol. 12, 928–937 (1995).
 Duret, L., Mouchiroud, D. & Gouy, M. Nucleic Acids Res. 22, 2360–2365 (1994).
 Novacek, M. Nature 356, 121–125 (1992).

- Higgins, D. G., Bleasby, A. J. & Fuchs, R. Comput. Appl. Biosci. 8, 189–191 (1992).
 Kumar, S., Tamura, K. & Nei, M. Molecular Evolutionary Genetics Analysis version 1.01
- (Pennsylvania State University, 1993)
- 22. Tajima, F. Genetics 135, 599-607 (1993)
- 23. Ota, T. & Nei, M. J. molec. Evol. **38**, 642–643 (1994).
- 24. Ramirez, V., Savoie, P. & Morais, R. J. molec. Evol. 37, 296-310 (1993).
- Xu. X. & Arnason, U. Gene 148, 357–362 (1994).
- 26. Takezaki, N., Rzhetsky, A. & Nei, M. Molec. Biol. Evol. 12, 823-833 (1995).
- 27. Hedges, S. B. & Sibley, C. G. Proc. natn. Acad. Sci. U.S.A. 91, 9861-9865 (1994).
- 28. Felsenstein, J. Phylogenetic Inference Package (PHYLIP), Version 3.5 (University of Washington,
- 29. Sibley, C. & Ahlquist, J. Phylogeny and Classification of Birds (Yale Univ. Press, New Haven, CT, 1990).
- 30. Smith, A. G., Smith, D. G. & Funnell, B. M. Atlas of Mesozoic and Cenozoic Coastlines (Cambridge Univ. Press, 1994).

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Correlation between male song repertoire, extra-pair paternity and offspring survival in the great reed warbler

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In many birds, females copulate with males other than their social mate, resulting in extra-pair fertilizations (EPFs)¹⁻⁷. It is still unknown, however, why females seek EPFs^{7,8}. In one study, males that accounted for most EPFs had higher survival⁶, but neither the characteristics revealing male quality nor the benefits accruing to females selecting attractive males were identified. Great reed warblers, Acrocephalus arundinaceus, are socially polygynous, and females base their mate choice on territory quality⁹ and song-repertoire size¹⁰, both of which predict harem size and reproductive success^{11,12}. By DNA fingerprinting¹³, we demonstrate that female great reed warblers obtain EPFs from neighbouring males with larger song repertoires than their social mate. In addition, the relative post-fledging survival of offspring was positively correlated with their genetical fathers' song repertoire size. These data support the hypothesis that females, by engaging in extra-pair fertilizations, seek genetic benefits for their offspring^{7,8}.

We studied a population of individually colour-ringed great reed warblers, a long-distance migrant with sexually monomorphic cryptic plumage, at lake Kvismaren, Sweden 9,12,14-18 from 1987 to 1993. All territories were visited daily throughout the breeding season (May–July). In this population, a substantial proportion of the returning migrants settle in their natal area¹⁸. Polygynous males (40% of territorial males^{12,14}) attract females sequentially about every 8 days¹⁶. Males have a loud mate-attraction

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